High-throughput microgel biofabrication via air-assisted co-axial jetting for cell encapsulation, 3D bioprinting, and scaffolding applications

Vaibhav Pal^{1,2\$}, Yogendra Pratap Singh^{2,3\$}, Deepak Gupta^{2,3}, Mecit Altan Alioglu^{2,3}, Momoka Nagamine^{1,2}, Myoung Hwan Kim^{2,4}, Ibrahim T. Ozbolat^{2,3,4,5,6,7,8*}

¹Department of Chemistry, Pennsylvania State University, University Park, PA, USA
²The Huck Institutes of the Life Sciences, Pennsylvania State University, University Park, PA, USA
³Engineering Science and Mechanics Department, Penn State University, University Park, PA, USA
⁴Department of Biomedical Engineering, Pennsylvania State University, University Park, PA, USA
⁵Materials Research Institute, Pennsylvania State College of Medicine, Hershey, PA, USA
⁶Department of Neurosurgery, Pennsylvania State University, Hershey, PA 17033, USA
⁸Department of Medical Oncology, Cukurova University, Adana, 01130, Turkey
*Corresponding author: Ibrahim T. Ozbolat. Email: ito1@psu.edu

Microgels have recently received widespread attention for their applications in a wide array of domains such as tissue engineering, regenerative medicine, and cell and tissue transplantation because of their properties like injectability, modularity, porosity, and the ability to be customized in terms of size, form, and mechanical properties. However, it is still challenging to mass (highthroughput) produce microgels with diverse sizes and tunable properties. Herein, we utilized an air-assisted co-axial device (ACAD) to continuously produce microgels in a high-throughput manner. To test its robustness, microgels of multiple hydrogels and their combination, including alginate (Alg), gelatin methacrylate (GelMA) and Alg-GelMA, were formed at a maximum production rate of ~65 000 microgels s⁻¹ while retaining circularity and a size range of 50–500 µm based on varying air pressure levels. The ACAD platform allowed single and multiple cell encapsulation with $74 \pm 6\%$ efficiency. These microgels illustrated appealing rheological properties such as yield stress, viscosity, and shear modulus for bioprinting applications. Specifically, Alg microgels have the potential to be used as a sacrificial support bath while GelMA microgels have potential for direct extrusion both on their own or when loaded in a bulk GelMA hydrogel. Generated microgels showed high cell viability (>90%) and proliferation of MDA-MB-231 and human dermal fibroblasts over seven days in both encapsulation and scaffolding applications, particularly for GelMA microgels. The developed strategy provides a facile and rapid approach without any complex or expensive consumables and accessories for scalable highthroughput microgel production for cell therapy, tissue regeneration and 3D bioprinting applications.